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Diagnosis, Prevention and Control Strategies of Infectious Bronchitis Virus

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Abstract

Avian infectious bronchitis (IB) is one of the illnesses related to respiratory disorders in household poultry causing critical financial misfortune to poultry yield. The IB can cause respiratory sickness, lessened creation of flying brutes, nephrotic disorder, and irredeemable hurt to the uterus, which can cause the odd curvature of eggs. Immunization programs are futile right now because of the excessive innate diversity of infectious bronchitis virus (IBV). Subsequently, a correct and quick serovar affirmation may be an important aspect to overcome IBV. Engrossing expository tools are moreover anticipated to diagnose IB illnesses within the area and to recognize different serovar and diversities. Perfect organization for hindering IB in flying brutes consolidates perfect vaccination with live or inactivated antibodies comprising streaming strains and serious disengagement of the sullied fowls. Extraordinary organization and sterile homes in poultry units can handle the escalation of IB among bunches. The thought of DNA immunization with spike protein quality has modified the idea of IB safe prophylaxis because it has been found to bring out a satisfactory immune response. Nuclear science-based distinguishing proof and control methods must be made nearby the up degree of standard procedures to handle the evolving threat displayed by this microorganism and the affliction can be satisfactorily controlled within the years.





Introduction

Infectious bronchitis (IB) infection is graded as avian coronavirus, which belongs to class Gammacoronavirus family Coronaviridae [1, 2]. It could be a familiar and broadly evolving infection, which causes huge financial misfortunes to the poultry industry by appearing respiratory and profitable regenerative signs, diminished exhibitions, and expanded mortality, especially when nephron pathogenic strains or auxiliary contamination is included [3]. Infectious bronchitis virus (IBV) is an encompassed infection with a stranded positive-sense RNA genome of approximately 27 kb, which depicts chromosome sequencing as follows: 5'UTR-1a/1ab-S-3b-3ba-E-M-5b-5a-N-3'UTR [4]. Infectious bronchitis could be an infection of financial significance influencing all the major poultry segments. It is stemmed from the United States influencing youthful chickens in 1931. Strains of the IBV infection cause transformation, which gets to be more deadly [5]. Whereas others stay constrained to a region with no slant for expansion [6]. In nations where the sick animal is taken after the contamination rate can reach up to 100% [7]. To halt the spread of IBV, it can be handled through live and dormant immunizations. Both sorts of antibodies have their downsides. The inert immunizations can merely enact the humoral resistant reaction, whereas no cellular reaction is actuated, it too improves the movement of lymphocytes [8, cytotoxic T 6]. Live immunizations can effortlessly tweak chromosome, which influences the spicules and in return diminishes.

After involving a few duplications cycles, the infection is hereditarily altered to "Quasispecies", particularly on their subunit S1 [9]. Active antibodies can increase the ratio of transformation up to 1.5% [10] and non-fulfillment of immunization can cause the evolution of the transformation [11]. The setting up of modern antibody procedures is critical for way better control of the malady [12]. Later inquiries demonstrated that recombinant immunizations are better than other customary immunizations. Diverse bacterial and viral specialists can be adjusted hereditarily to act as vectors that appear diverse qualities encoding the major basic viral proteins. The defensive insusceptibility is created when we utilize recombinant antibodies. The substitutes and suitable choices are based on total

information of the disease and defense response nature, which offer assistance to choose antigen [13]. The core of this research is to elaborate the infinitesimal characteristics of chicken IBV, to expound the cellular resistant and humoral reactions, particularly those played by cytotoxic T lymphocytes and partly played by spike (S) nucleoprotein (N) and glycoprotein (S) in the acceptance of resistant reaction. Hereditary treatment and other biotechnological progress in infectious bronchitis control have been evaluated by numerous modern researchers.

Diagnosis

Identification of IB can be performed by the clinical examination of immunoglobulin titers. disease activation signs, and acclaimed DNA of IBV tissues from a respiratory organs-like pulmonic, kidney and windpipe of debilitated brutes [14-16]. The location of antigen diagnosis is the liver and pancreas while spleen and windpipe are for scanning or microscope-based scanning of tissues [17]. Detachment of the essential infection requires two to three dazzle sections, which may be troublesome and lengthy, for that the fetus of hen is unconfined with specific organisms and their organ cultures of trachea (OCT) are suggested for division of IBV [18]. Other identification strategies like histochemistry [19, 20] or in situ assimilation can be utilized [21]. We employ a magnifying instrument to identify the protein (N) from healthy and safe cells, it indicates the presence of viral proteins (N), which is exceptionally vital for duplication in the hold [22]. Moreover, we can identify IB by diverse tests like virus neutralization test (hemagglutination inhibition),

Prevention and Control Strategies

Immunological control (Vaccination)

As a routine practice in many countries, day-old chicks are passed through the process of vaccination, which enhances their immunity against IBV. Mostly, the chicks are being vaccinated through the water as it is the easiest and most economical way of vaccination without any labor charges. We must use a vaccine of lower virulence as a higher level of virulence causes respiratory reactions. As a result, signs are visible as the immunity of the chick is too low that cannot

protect the respiratory tract [23]. There are three types of vaccines: live, inactivated and killed vaccines. Live vaccines are usually used as broiler, breeder and layer's primary vaccine in commercial farming. A vaccine is usually injected when the layer or breeder is near to period of laying eggs [24]. Vaccination helps to boost immunity for a long period thus, this would provide long life immunity [25].

Live vaccine

Massachusetts strain H-120 is the common example of live vaccine and it is a classical and effective vaccine against the disease. It is an important vaccine at the start of the flock, which produces long-term immunity without having any bad impact on a bird's immune system [26]. Initially, the Immunization would be done via the ocular, intramuscular route, or tracheal route. This method is most economical; increases regional and whole systematic immune response. But there is a possibility of sudden vaccine reactions after a few days of vaccination [27]. Ma5, a single segment immunization, is not severe, a single segment can be added in the first program of immunization with IB 4/91 vaccines and inactivated immunization provide immunity from many IB virus variants. Live immunization is normally practiced on layers and breeders at a young age to keep the regional protection of respiration tract and is recommended in the location of ultra-level of field challenges. However, vaccine strain should be selected based on strains prevalent in local areas and countryside farming. Protection against analogous, reference strains and serotype fields can be provided by the vaccine [28]. The occurrence of different variants of the virus has very complex processes and increases disease prevention costs. It is recommended to use the regional strains in vaccines for efficient protection against disease. The function of activated immunization is to avoid depression and decrease the number of eggs and meat production. The Massachusetts (Mass or M41) strain is the most well-known because it represents initial isolates from many areas [29, 30].

Inactivated vaccines

Inactivated vaccines initiate enduring insusceptibility, show no inoculation responses, normally cost more than live immunizations and blend of various antigens separated from IBV can

be accomplished when given exclusively. The raised degree of flowing antibodies was animated by inactivated immunizations than live sera; in this manner, it is valuable in a reproducer program where maternal counter antibody security is required. Still, because of enlistment of better T cell reactions and delivering a higher immune response (IgA) feeling, changed live antibodies assume a critical part in ensuring business birds (layers). Chickens should be appropriately prepared with live immunization to take better advantage of the inactivated immunizations, and by this way, most elevated titers will be acquired in a period of four and half months (time of inoculation) between the last live and inactivated antibody [31]. Further immunization projects might be rearranged by consolidating inactivated antigens against at least two serotypes (or at least two infections) into one antibody [32].

Environmental control

The control is highly dependent on good management practices by proper bird density in the farm, quality of air, following strict biosecurity measures, etc. [33]. However, even favorably handled IBV-positive flocks were estimated to yield 3% less than the free flock of IBV [34, 35]. The first barrier of IBV is to follow strict biosecurity measures [36, 37]. Disinfection of the farm is one of the major tools in minimizing the risks associated with the IBV virus. However, biosecurity alone has a rare chance of completely preventing disease transmission. Vaccination of the flock should be done according to the vaccination protocol. Still, it cannot fully assure the prevention but decrease in the incidence of disease in the flock. For instance, H120-vaccinated animals showed a reduced viral transmission (R0<1) and shedding after homologous challenge in the condition of experiments [38].

Future Perspectives

With the advances of microbiology in different laboratories, novel vaccines like subunit, vector, and DNA vaccine using glycoprotein (S1) gene along with the reserve genetic vaccine have been tried [39]. The utilization of DNA (S protein-based DNA) antibodies changes the idea of inoculation as opposed to infectious bronchitis [16]. Alike, customized antibodies would be intended to suit the locally overall infectious bronchitis virus strains and the issue of weakened live strains

returning to harmfulness can stay away. The vector-based or recombinant antibodies are additionally intended to present antigens from at least two infections, which go about as multivalent immunization assuring at least a couple of illnesses. DNA immunization is another promising zone because appeared in starting clinical preliminaries. Such new-age immunizations could be managed securely in eggs or chickens. The adequacy of such immunizations could be tried in huge scope tests before present in order for a business reason [40, 41]. This requires great importance to battle monetarily significance and arising poultry microorganisms by adjusting and creating more current diagnoses, compelling more secure antibodies, investigating therapeutics, and fitting avoidance and handling procedures.

Conclusions

This review shows how we can detect infectious bronchitis virus and how we can fix it. Moreover, this review summarizes the modern knowledge of IBV. The results of modern Vaccinology genetic research are very motivating and promising and the control of IBV has recorded a great evolution with recombinant vaccines.

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Conflict of Interest

The authors declare that they have no conflict of interest

References

- [1] Ismael II, Hasan, Rasheed TS, Jasim NA, Shakor MK. Pathological effect of infectious bronchitis disease virus on broiler chicken trachea and kidney tissue. Vet World 2020; 13(10):2203-2208.
- [2] Woo PCY, Lau SKP, Lam CSF, Lau CCY, Tsang AKL, Lau JHN, et al. Discovery of seven novel mammalian and avian coronaviruses in the genus Delta coronavirus supports Bat coronaviruses as the gene source of alphacoronavirusand betacoronavirusand avian coronaviruses as the gene source of

- gammacoronavirus and deltacoronavirus. J Virol 2012; 86:3995-4008.
- [3] Franzo G, Legnardi M, Tucciarone CM. Evolution of infectious bronchitis virus in the field after homologous vaccination introduction. Vet Res 2019; 50(1):92.
- [4] Legnardi M, Tucciarone CM, Franzo G, Cecchinato M. Infectious bronchitis virus evolution, diagnosis and control. Vet Sci 2020; 7(2):79.
- [5] Rafiei MM, Vasfi-Marandi M, Bozorgmehri-Fard MH, Ghadi S. Identification of different serotypes of infectious bronchitis viruses in allantoic fluid samples with single and multiplex RT-PCR. J Virol 2010; 3:24-29.
- [6] Wu SW, Li L, Wang Y, Xiao Z. CTL-derived exosomes enhance the activation of CTLs stimulated by lowaffinity peptides. Front Immunol 2019; 10:1274.
- [7] Barry J, Goudar MS, Nighot PK, Kshirsagar SG, Ladman BS. Emergence of a nephron pathogenic avian infectious bronchitis virus with a novel Genotype in India. J Clinic Microbiol 2005; 43:916-918.
- [8] Cavanagh D Coronavirus avian infectious bronchitis virus. Vet. Res 2007; 38: 281-297.
- [9] Liu S, Zhang X, Gong L, Yan B, Li C. Altered pathogenicity, immunogenicity, tissue tropism and 3'-7 kb region sequence of an avian infectious bronchitis Coronavirus strain after serial passage in embryos. Vaccine 2009; 27:4630-4640
- [10] Lee CW, Jackwood MW. Origin and evolution of Georgia 98 (GA98), a new serotype of avian infectious bronchitis virus. Virus Res 2001; 80:33-39.
- [11] Cavanagh D, Davis PJ, Cook JK, Li D, Kant A, Koch G. Location of the amino acid differences in the S1 spike glycoprotein subunit of closely related serotypes of infectious bronchitis virus. Avian Pathol 1992; 21:33-43.
- [12] Yan F, Zhao Y, Hu Y, Qiu J, Lei W. Protection of chickens against infectious bronchitis virus with a multivalent DNA vaccine and boosting with an inactivated vaccine. Vet Sci 2013; 14:53-60.
- [13] Nascimento P, Leite LCC. Recombinant vaccines and the development of new Vaccine strategies. Braz J Med Biol Res 2012; 45: 1102-1111.
- [14] Pradhan HK, Mohanty GC, Verma KC. Isolation and characterization of viral agents from the reproductive tract of young chicks Indian J Poult Sci 1982; 17:143-148
- [15] Verma KC, Malik BS. Isolation of infectious bronchitis virus of poultry in India. Indian Vet J 1971; 48:887-892.
- [16] Sylvester SA, Kataria JM, Dhama K, Senthilkumar N, Bhardwaj N, Rahul S. Detection of avian infectious bronchitis virus infected allantoic fluid using SI gene serotype specific RT-PCR. Indian J Comp Microbiol Immuno Infect Dis 2003; 24:39-42.
- [17] Fan WQ, Wang HN, Zhang Y, Guan T, Wang ZB. Comparative dynamic distribution of avian infectious Bronchitis virus M41, HI 20 and SAIBK strains by quantitative real-time RT-PCR in SPF chickens. Biosci Biotechnol Biochem 2013; 76:2255-2260.

- [18] Cook JK, Darbyshire JH, Peters RW. The use of chicken tracheal organ cultures for the isolation and assay of avian infectious bronchitis virus. Arch. Virol 1976; 50:109-118.
- [19] Nakamura K, Cook JK, Otsuki K, Huggins MB, Frazier JA. Comparative study of respiratory lesions in two chicken lines of different susceptibility infected with infectious bronchitis virus: histology, ultrastructure and immunohistochemistry. Avian Pathol 1991; 20:241 -257.
- [20] Chen BY, Hosi S, Nunoya T, Itakura C. Histopathology and immunohistochemistry of renal lesions due to infectious bronchitis virus in chicks. Avian Pathol 1996; 25:269-283.
- [21] Collison S EW, Li J, Sneed LW, Peters ML, Wang L. Detection of avian infectious bronchitis viral infection using in situ hybridization and recombinant DNA. Vet Microbiol 1990; 24:261-271.
- [22] Hiscox JA, Wurm T, Wilson L, Britton P, Cavanagh D, Brooks G. The coronavirus infectious Bronchitis virus nucleoprotein localizes to the nucleolus. J Virol 2001; 75:506-512.
- [23] Kataria JM, Madan MC, Sohini D, Dash BB, Dhama K. Diagnosis and immune prophylaxis of economically important poultry diseases: A reviewIndian J Anim Sci 2005; 75:555-567.
- [24] Jackwood MW, Hilt DA, Callison SA. Detection of infectious bronchitis virus by real-time reverse transcriptase-polymerase chain reaction and identification of a quasispecies the Beaudette strain. Avian Dis 2003; 47:18-724.
- [25] Cook JK, Orbell SJ, Woods MA, Huggins MB. Breadth of protection of the respiratory tract provided by different live-attenuated infectious bronchitis vaccines against challenge with infectious bronchitis viruses of heterologous serotypes. Avian Pathol 1999; 28:477-485.
- [26] Matthijs MG, van Eck JH, Landman WJ, Stegeman JA. Ability of Massachusetts-type infectious Bronchitis virus to increase colibacillosis susceptibility in commercial broilers: A comparison between vaccine and virulent field virus. Avian Pathol 2003; 32:473-481.
- [27] Bijlenga G, Cook JKA, Gelb J, de Wit JJ. Development and use of the Hstrain of avian infectious Bronchitis virus fromthe Netherlands as a vaccine: A review. Avian Pathol 2004; 33:550-557.
- [28] de Wit JJ, van de Sande H. Efficacy of combined vaccines at day of hatch against a D388 challenge in SPF and commercial chickens. Proceedings of the 6th International Symposium on Corona and Pneumoviruses and Complicating Pathogens June14-17 Rauischholz hausen Germany 2009; pp. 177-182.
- [29] Gelb J, Wolff JB and Moran CA. Variant serotypes of infectious bronchitis virus isolated from commercial layer and broiler chickens. Avian Dis 1991; 35:82-87.

- [30] Terregino C, Toffan A, Beato MS, De Nardi R, Vascellari M. Pathogenicity of a QX strain of infectious Bronchitis virus in specific pathogen free and commercial broiler chickens and evaluation of protection induced by a vaccination programmed based on the Ma5 and 4/91 serotypes. Avian Pathol 2008; 37:487-493.
- [31] Ladman BS, Pope CR, Ziegler AF, Swieczkowski T, Callahan CJ, Davison S, Gelb Jr J. Protection ofchickensafterliveandinactivatedvirusvaccinatio nagainstchallengewith nephron pathogenic infectious bronchitis virus PA/Wolgemuth/98.Avian Dis 2002; 46:938-944.
- [32] Hong SM, Kwon HJ, Kim IH, Mo ML Kim JH. Comparative genomics of Korean Infectious Bronchitis viruses (IBVs) and an animal model to evaluate pathogenicity of IBVs to the reproductive organs. Viruses 2012; 4:2670-2683
- [33] Ignjatovic J, Sapats S. Avian infectious bronchitis virus. Rev Sci Tech 2000; 19:493–508.
- [34] van Ginkel FW, Padgett J, Martinez-Romero G, Miller MS, Joiner KS, Gulley SL. Age-dependent immune responses and immune protection after avian coronavirus vaccination. Vaccine 2015; 33:2655– 2661
- [35] McMartin DA. Virus Infections of Vertebrates. Virus Infections of Birds. Volume 4. Elsevier Science Publishers; Amsterdam, The Netherlands; 1993, pp. 249–275.
- [36] Racicot M, Vaillancourt JP. Biosecurity: Assessing and Managing Risks; Proceedings of the 12e Journées de La Recherche Avicole et Palmipèdes à Foie Gras; Tours, France 2017; pp. 20–34.
- [37] van Limbergen T, Dewulf J, Klinkenberg M, Ducatelle R, Gelaude P, Méndez J, et al. Scoring biosecurity in European conventional broiler production. Poult Sci 2018; 97:74–83.
- [38] de Wit JJ, de Jong MC, Pijpers A, Verheijden JH. Transmission of infectious bronchitis virus within vaccinated and unvaccinated groups of chickens. Avian Pathol 1998; 27:464-471.
- [39] Dhama K, Mahendran M. Technologies and advances in diagnosis and control of poultry diseases: Safeguarding poultry health and productivity Poult Technol 2008; 2:13-16.
- [40] Boots AM, Benaissa-Trouw BJ, Hesselink W, Rijke E, Schrier C, Hensen EJ. Induction of anti-viral immune responses by immunization with recombinant-DNA encoded avian coronavirus nucleocapsid protein. Vaccine 1992; 10:119-124.
- [41] Yu LZ, Wang, Jiang Y, Low S, Wang JK. Molecular epidemiology of infectious bronchitis virus isolates from China and Southeast Asia. Avian Dis 2001; 45:201-209.